

Protein and Ascorbic Acid Changes in Shoot of Certain C₃ and C₄ Plants in Relation to Light and Dark

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(Received 3 February 1981)

Changes in soluble protein and ascorbic acid contents have been analysed at various growth stages in shoot of *Hordeum vulgare* L., *Lathyrus odoratus* L. (C₃ plants); *Sorghum vulgare* Pers. and *Zea mays* L. (C₄ plants). The quantity of ascorbic acid increased with the growth of seedlings and significantly higher concentration of this metabolite as well as protein in the shoot at flowering stage in comparison to 28-days-old vegetative stage in all the plants may indicate increased requirement for the highly-activated stage of differentiation. The amounts of protein and ascorbic acid were also found significantly higher in light than in dark-grown seedlings. Interestingly, loss of ascorbic acid in C₄ plants was significantly less than that in C₃ plants maintained in dark. Moreover, C₄ plants were found more efficient in regaining the lost ascorbic acid level on exposure to light in comparison to the C₃ plants.

Key Words: Protein, Ascorbic acid, C₃ plants, C₄ plants

Introduction

The role of ascorbic acid in germination, seedling growth, differentiation, flowering and drought resistance has been extensively studied and reviewed (Mapson 1958, Isherwood & Mapson 1962, Chinoy et al. 1971). Ascorbic acid metabolism in relation to proteins and nucleic acids was also investigated with the onset of flowering (Chinoy 1962, 1964, Chinoy et al. 1973). Since, ascorbic acid is known to have diverse functions (Mapson 1958); the present work was chiefly aimed to compare certain C₃ and C₄ plants for the

total soluble protein and ascorbic acid contents of shoot at different growth stages in light and dark.

Materials and Methods

Seeds of two C₃ plants viz., *Hordeum vulgare* L., *Lathyrus odoratus* L. and two C₄ plants viz., *Sorghum vulgare* Pers. and *Zea mays* L. were selected and surface sterilized with 0.01% mercuric chloride for 1 min. These seeds, after 16 hr of water soaking (designated as 'initial' stage) were sown on absorbent paper moistened with

distilled water in Petridishes, For analysis, the seedlings were harvested after 48 and 96 hr from the 'initial' stage. The seedlings were grown in water culture upto 96 hr under (i) a fluorescent tube light (970 lux) and (ii) complete darkness. After 96 hr stage, the seedlings were transferred in earthen pots (45 × 30 cm), each containing 2.5 kg of garden soil (pH=7.8) and 2 kg of dung manure. The pots were kept continuously in natural light and watered every third day. Four replicates of samples from the potted plants were taken 7, 14 and 28 days, after the 'initial' stage, and lastly at the initiation of flowering. At 28-days stage, plants were kept in dark for 48 hr and then exposed to natural light for 0.5, 1.0, 2.0 and 4.0 hr. Shoot (3rd nodal leaf from apex) samples were taken for analysis at these stages.

The protein estimation was done following Lowry et al. (1951) using Folin's reagent. The ascorbic acid was extracted following Bessey and King (1933). Five-hundred mg of plant tissue was crushed in 25 ml of 5% metaphosphoric acid and digested on a water bath for 10 min. The extract was cooled, filtered and final volume was raised to a specific volume. Seven ml of this filtrate was titrated against 2, 6-dichlorophenol indophenol solution until a distinct pink-rose colour persisted at least for 5 sec.

The ascorbic acid content of the extract was calculated by the formula (AOAC 1970):

$$\text{Ascorbic acid (mg)/100 g tissue} \\ = I \times S \times D / A \times 100 / W$$

where I =Indophenol reagent used in titration (ml); S =Ascorbic acid reacting with 1 ml of the reagent (mg); D =volume of the extract (ml); A =aliquot titrated (ml); W =weight of the sample (g)

Results and Discussion

Protein and ascorbic acid changes at various growth stages in all the plants have been presented in table 1. The data at these stages were statistically analysed using 't' test and the results were found to be statistically significant at $P > 0.05$

The quantity of ascorbic acid in the shoot increased with the growth of seedlings upto flowering stage except a decline at 28-days-old stage in the C_4 plants. However, the decrease was significant only in *Z. mays*. The shoot at flowering stage had significantly higher concentration of protein as well as of ascorbic acid than in the vegetative shoot of 28-days-old stage in all the plants. The production and utilization of ascorbic acid at higher levels during the period of reproductive differentiation produce a highly-activated metabolic state in the cell due to which proteins and enzymes are synthesized at faster rates (Chinoy 1962, 1964).

A perusal of results showed that the amount of protein and ascorbic acid was significantly higher in light-grown than in the dark-grown seedlings in almost all the comparable samples studied. It has been established earlier also that protein content is higher in light-grown and low in dark-grown seedlings (Rai & Laloraya 1965, 1967, Mukherjee & Laloraya 1979).

The amount of ascorbic acid as well as of protein decreased in all the plants when kept in dark for 48 hr at 28-days-old stage. However, the decline in ascorbic acid content was more prominent in C_3 plants than in C_4 plants. Interestingly, ascorbic acid content in C_4 plants was either equal or more than the initial level of 28-days-old stage, when the final data of 4.0 hr-light stage was compared, C_3 plants, however, failed to regain it. On comparing the levels of ascorbic acid at 48 hr dark and 4.0 hr light stages, the maximum increase in light could be noticed in *Z. mays*

Table 1 Mean values of total soluble proteins (mg/g of dry wt) and ascorbic acid (mg/100 g fr. wt) of shoot at various growth stages in certain C₃ and C₄ plants

Growth stages	Protein						Ascorbic acid					
	C ₃ plants			C ₄ plants			C ₃ plants			C ₄ plants		
	<i>H. vulgare</i>	<i>L. odoratus</i>	<i>S. vulgare</i>	<i>H. vulgare</i>	<i>L. odoratus</i>	<i>Z. mays</i>	<i>H. vulgare</i>	<i>L. odoratus</i>	<i>S. vulgare</i>	<i>H. vulgare</i>	<i>L. odoratus</i>	<i>Z. mays</i>
*48 hr	36.00	—	90.64	44.00	—	42.56	44.00	—	28.40	—	32.80	
96 hr	24.90	85.31	120.60	71.00	106.50	78.50	71.00	106.50	47.85	51.75	51.75	
48 hr	79.73	—	102.80	51.50	—	49.71	51.50	—	35.60	—	50.44	
96 hr	74.30	119.97	127.00	80.00	142.00	123.70	80.00	142.00	56.76	64.55	64.55	
7 days	149.95	141.20	66.51	85.00	148.00	90.65	85.00	148.00	58.00	66.10	66.10	
14 days	81.90	139.55	45.50	95.00	149.50	34.99	95.00	149.50	62.55	70.00	70.00	
28 days	56.34	106.70	28.14	98.00	152.00	44.80	98.00	152.00	65.60	52.78	52.78	
(i) 48 hr	52.10	95.33	26.18	65.00	98.50	40.40	65.00	98.50	63.50	42.56	42.56	
(ii) 0.5 hr	60.50	124.66	24.80	68.50	102.00	38.70	68.50	102.00	60.46	54.50	54.50	
(iii) 1.0 hr	109.00	103.88	25.50	70.50	101.00	43.20	70.50	101.00	64.55	55.45	55.45	
(iv) 2.0 hr	96.95	97.96	24.40	74.00	120.50	35.47	74.00	120.50	68.00	57.42	57.42	
(v) 4.0 hr	87.70	76.40	23.00	75.00	126.00	32.85	75.00	126.00	70.60	60.55	60.55	
Flowering stage	119.50	125.50	65.80	111.50	198.50	86.60	111.50	198.50	74.65	77.80	77.80	

* Shoot did not appear at 48 hr stage in *L. odoratus*

followed by *L. odoratus*, *H. vulgare* and *S. vulgare*. Overall results of ascorbic acid changes may indicate that its minimum loss in C₄ plants than in C₃ plants is equal or more effective than its synthesis in light in regaining the level of 28 days at 4.0 hr-light stage. Chinoy (1966) had pointed out earlier that during increased production of ascorbic acid, the carbon cycle also contributed substantially and the process of photophosphorylation and oxidative phosphorylation were further energized by the flow of electrons from monodehydroascorbic acid (MDHA) -the product of ascorbic acid oxidation.

Thus, it appears that to meet the higher pool of ATP in C₄ plants, ascorbic acid is being synthesized rapidly and it gets oxidized to a powerful reducing agent-MDHA, which further activated the phosphorylation processes.

Acknowledgements

Authors are grateful to Professor R S Mehrotra, Head, Department of Botany for providing laboratory facilities and to the Council of Scientific and Industrial Research, for financial assistance to one of us (B S A).

References

- AOAC 1970 in *Official Methods of Analysis*; ed. W Horwitz (Washington: Assoc. of Official Anal. Chem.) 769 pp
- Bessey O A and King G G 1933 The distribution of vitamin C in plant and animal tissues and its determination; *J. Biol. Chem.* 103 687-698
- Chinoy J J 1962 Formation and utilization of ascorbic acid in the shoot apex of wheat as factors of growth and development; *Indian J. Pl. Physiol.* 5 172-195
- 1964 Ascorbic acid-nucleic acid-protein metabolism. Concept of flowering in plants. *Proc. 10th Intl Bot. Congr. Absts.* p 347
- 1966 Role of correlations between growth, metabolism and development in elucidating the mechanism of their heredity. *J. Indian bot. Soc.* 45 150-174
- , Singh Y D and Gurumurti K 1971 Some aspects of the physiological role of ascorbic acid in plants, *Indian Agric.* 15 33-48
- , Shah C K and Suthar H K 1973 Changes in ascorbic acid content in the shoot apex during reproductive differentiation in maize; *Indian J. Pl. Physiol.* 16 7-15
- Isherwood F A and Mapson L W 1962 Ascorbic acid metabolism in plants. II. Biosynthesis; *Ann. Rev. Pl. Physiol.* 13 329-350
- Lowry O H, Rosebrough N J, Farr A L and Randall R J 1951 Protein measurement with the folin phenol reagent; *J. Biol. Chem.* 193 265-275
- Mapson L W 1958 Metabolism of ascorbic acid in plants: Part I. Function; *Ann. Rev. Pl. Physiol.* 9 119-150
- Mukherjee D and Laloraya M M 1979 Nitrogen and free amino acid changes during seedling growth in *Bauhinia purpurea* J. *Indian. bot. Soc.* 58 75-82
- Rai V K and Laloraya M M 1965 Correlative studies on plant growth and metabolism. I. Changes in protein and soluble nitrogen accompanying gibberellin-induced growth in lettuce seedlings; *Pl. Physiol* 40 437-441
- and — 1967 Correlative studies on plant growth and metabolism. II. Effect of light and gibberellic acid on the changes in protein and soluble nitrogen in lettuce seedlings; *Pl. Physiol.* 42 440-444