

*Review Article***Cell-Derived Microparticles (MPs) and Their Role in Unexplained Recurrent Pregnancy Loss**

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(Received on 31 January 2018; Revised on 14 June 2018; Accepted on 18 June 2018)

Circulating cell-derived microparticles (MPs) are sub-micronic phospholipid vesicles derived from cells of different types in response to various biological processes such as cellular activation and apoptosis. They are mainly biomarkers of activation, damage and stress. Apart from the pro-thrombotic attribute, MPs have many other properties and various functions such as being pro-inflammatory, pro-angiogenic or immunomodulatory; and have been found to have a role in vascular dysfunction, cross talk, inflammation, etc. There are many studies which show the association of specific cell-derived MPs to different pathological states. MPs act as possible predictors of disease severity and the origin of MPs help us predict the pathological condition; thus MPs represent promising biomarkers of the disease state. There are several methods for the detection and characterization of MPs; however the most common and gold standard method is flow cytometry. Other methods include enzyme-linked immunosorbent assay and functional assays which measure procoagulant activity and techniques such as electron microscopy, atomic force microscopy, nanoparticle tracking. However, research in the area of MPs faces many problems with regards to the methodology and pre-analytical variables and thus strict guidelines need to be followed for MP estimation. Elevated procoagulant MPs have been detected in patients with pregnancy complications and unexplained recurrent pregnancy loss. MPs may assist in uteroplacental thrombosis leading to pregnancy complications and miscarriages. A preliminary study reports the beneficial use of anticoagulant therapy in such patients with elevated MPs. Present report summarizes different methods used for the detection and analyses of MPs along with their role in pregnancy complications mainly unexplained recurrent pregnancy loss.

Keywords: Cell-Derived Microparticles; Thrombosis; Flow Cytometry; Pregnancy Loss; Anticoagulant Therapy

Introduction

Circulating cell-derived microparticles (MPs), once perceived as simply “innocent debris” and referred as “platelet dust” in the late sixties (Wolf, 1967; Boulanger *et al.*, 2011) is now emerging as a novel biomarker in a wide range of clinical settings. They are biomarkers reflecting cell activation and cell death; having many attributes such as being procoagulant, pro-inflammatory, etc. and play a major role in intercellular communication. They are emerging as predictive biomarkers for different clinical diseases; as well as circulating biologically active entity (Jy *et al.*, 2010). The research on MPs in different areas is growing at a very rapid pace. The methods for detecting and analyzing these MPs are ever evolving

with flow cytometry (FC) technique being the gold standard presently.

One of the hypothesized causes of unexplained and/ or recurrent pregnancy loss (RPL) and other pregnancy complications is hypoxia due to uteroplacental thrombosis (Rai, 2003). Nowadays, both genetic thrombophilia and antiphospholipid antibodies are being assessed for the same reason. But even after these investigations, many cases remain idiopathic. There are studies which now report elevated levels of procoagulant MPs in women with pregnancy complications which may contribute to thrombosis in the uteroplacental vasculature leading to placental insufficiency. If this holds true, the next question that arises is whether anticoagulant therapy (ACT) will prove to be beneficial in these cases.

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In this review, we have shed light on what are these cell-derived MPs; the different methods for MP assessment available, their role in unexplained RPL and the clinical implications.

What are Cell-derived Microparticles (MP)?

Circulating MPs are sub-micronic phospholipid vesicles, 0.1- 1 μm in size, derived from cells of different types in response to various biological processes such as cellular activation and apoptosis. MPs harbor a concentrated set of mRNA, proteins, microRNA, etc. and appear to transfer these components to nearby or remote cells (Garcia *et al.*, 2005; Ratajczak *et al.*, 2006; Hunter *et al.*, 2008; Gibbins *et al.*, 2009). Circulating MPs are found in healthy, normal state in a particular range specific to the parent cell from which they are derived; in the absence of disease these MPs originate from aging cells (Zubairova *et al.*, 2015).

The mechanism of MP formation involves budding of the outer cell membranes which shows cytoskeletal re-organization and the externalization of phosphatidylserine (PS) on the surface, bestowing MPs their procoagulant activity. Lipid rich microdomains have been implicated in endothelial, platelet and monocyte derived MP formation (Burger *et al.*, 2013). The mechanism of formation and size differentiates MPs from exosomes as shown in Fig. 1. Exosomes originate by the inward budding of the membrane which leads to the formation of multivesicular bodies (size: 30-100 nm).

Certain populations of MPs have also been shown to express tissue factor (TF) on their surface (Owens and Mackman, 2011; Lacroix and Dignat, 2012). TF expressing MPs are important for thrombin generation and thus play a vital role in blood clotting in vitro (Hrachovinová *et al.*, 2003) as well as thrombus formation in vivo (Aras *et al.*, 2004; Furie and Furie, 2005; Tesselaar *et al.*, 2007; Khorana *et al.*, 2008; Owens and Mackman, 2011; Van Der Meijden *et al.*, 2012) as shown in Fig. 2. PS expressing MPs are thus used as the main marker to identify and quantitate MPs especially in clinical settings associated with thrombosis.

Apart from the pro-thrombotic attribute, MPs have several properties and functions such as being pro-inflammatory, pro-angiogenic or immunomodulatory and have a role in vascular dysfunction, cross talk, inflammation, etc. (Jy *et al.*, 2010). These MPs contribute to pathological states through different mechanisms such as angiogenesis, inflammation and coagulation.

MPs are derived from different cells like endothelial cells, erythrocytes and leukocytes; however the maximum percentage of total MPs is represented by platelets or megakaryocytes (Horstman and Ahn, 1999; Flaumenhaft *et al.*, 2009; Burnier *et al.*, 2009). MPs derived from different cell types possess different functional capabilities due to variations in proteins and lipids derived from the parent cells (Jimenez *et al.*, 2003; Perez-Pujol *et al.*, 2007; Aleman *et al.*, 2011). There are many studies which

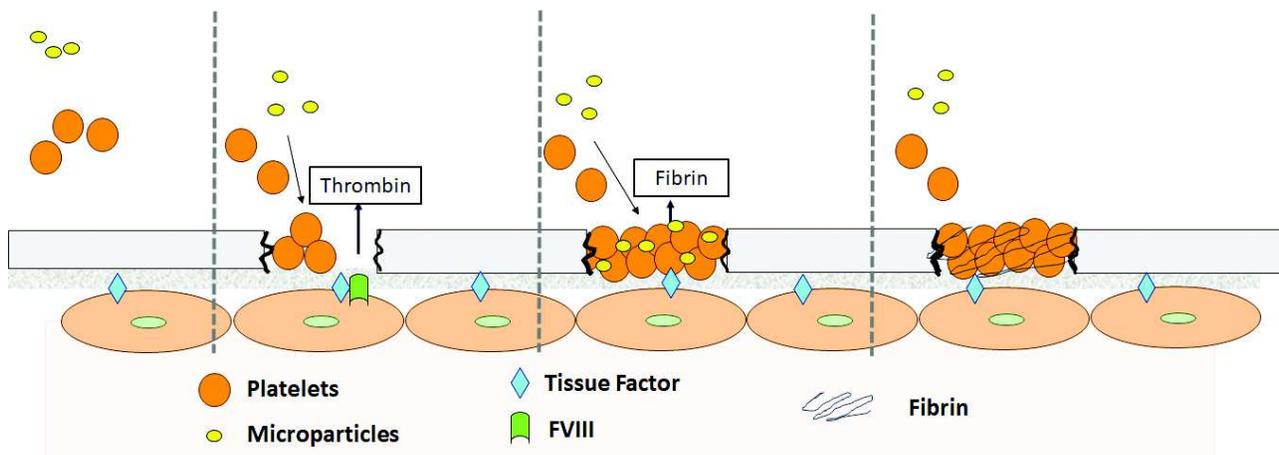


Fig. 1: Mechanism of formation of microparticles and exosomes a) Cell derived microparticles 100-1000 nm: MPs are formed by outward budding of the plasma membrane involves cytoskeletal re-organization and the externalization of phosphatidylserine on the surface, bestowing them their procoagulant activity. b) Exosomes 30- 100 nm: Inward budding of the membrane leads to multivesicular bodies; from which exosomes are released

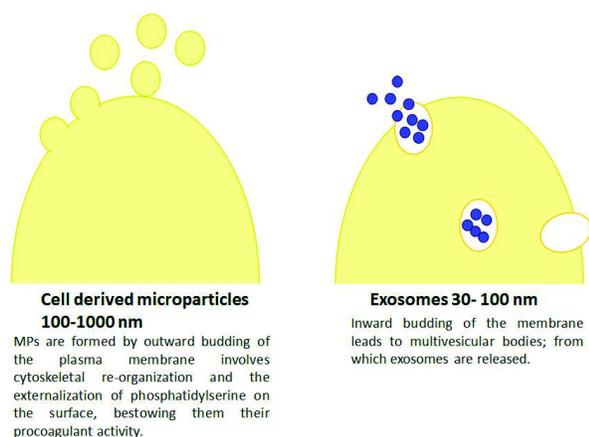


Fig. 2: Role of MPs in clot formation: MPs have a role in the formation of clot at the site of blood vessel injury. Once there is an injury to the blood vessel, platelets bind rapidly to the subendothelium and TF expressed on vessel cells activates and triggers the coagulation cascade and binds to FVIII to result in thrombin generation. MPs lead to the amplification of this coagulation cascade; after which fibrin clot is stabilized

have found an association between a particular clinical disorder and elevated MP levels of specific cell type.

However despite these studies on the role of MPs in different diseases, their role in healthy individuals is unclear. MPs are shown to support thrombin generation but whether those in circulation in healthy individuals can provide sufficient activity for the same is unclear (Berckmans *et al.*, 2001). One study showed that MPs have significant causative effects on fibrin polymerization and on the final structure of fibrin clot. MPs accelerate fibrin polymerization and support the formation of lysis resistant hemostatic fibrin clots as well as the thrombi and clots formed in clinical disorder. Therefore, the blood of healthy individuals contains functional MPs at the levels that have a procoagulant potential; they affect the structure and stability of fibrin clots indirectly through acceleration of thrombin generation and through direct physical incorporation into the fibrin network increasing resistance to fibrinolysis (Zubairova *et al.*, 2015).

Research on cell-derived MPs is mainly focused on their potential to act as biomarkers of coagulation, inflammation, endothelial dysfunction, and other pathological processes in different clinical settings to act as predictors. Elevated PS expressing and platelet

MPs (PMPs) is associated with a prothrombotic state, increase in endothelial MPs (EMPs) reflects vascular injury and elevated leucocyte MPs indicates a proinflammatory state. However, as many of these processes overlap, MPs of different type may be elevated or reduced in different pathologies (Burger *et al.*, 2013).

There are many studies which show the association of specific cell-derived MPs to different pathological states. Platelet, leucocyte and endothelial MPs are found elevated in diabetes (Omoto *et al.*, 1999; Omoto *et al.*, 2002; Koga *et al.*, 2005; Feng *et al.*, 2010; Tramontano *et al.*, 2010) and pulmonary hypertension (Amabile *et al.*, 2008, 2009; Diehl *et al.*, 2011). Elevated MP levels have been found in different thrombotic conditions (Zahra *et al.*, 2011) like atherosclerosis, pulmonary hypertension, heart failure and also end-stage kidney disease (Amabile *et al.*, 2005, 2012; Faure *et al.*, 2006); as well as in pregnancy complicated conditions like preeclampsia (PE) (Jadli *et al.*, 2017), intra uterine fetal death, RPL (Patil *et al.*, 2013) etc. Elevated red blood cell MPs have also been found associated with severe dengue (Punyadee *et al.*, 2015). Thus, MPs act as a possible predictor of disease severity; and their origin helps us predict the pathological condition making them promising biomarker of the disease state.

How are these Cell-Derived MPs Detected and Analyzed?

MPs are shed by all cell populations; however their measurement and detection are restricted to biological fluids. Majority of the studies analyze MP levels in circulating blood; while few have detected MPs in cerebrospinal fluid (Morel *et al.*, 2008), urine (Lytvyn *et al.*, 2017), lung fluids (Roca *et al.*, 2016), sputum (Lacedonia *et al.*, 2016), saliva (Suptawiwat *et al.*, 2017), etc. There are several methods of MP detection and characterization; however the most common method is FC (Flow cytometry). By FC method, one can not only gate the size of MPs but also characterize the cellular origin and quantitate their levels. It also helps in analysing large number of samples in one go. The other common methods of MP detection include enzyme linked immunosorbent assay (ELISA) and functional assays which measure procoagulant activity. Techniques like electron microscopy, atomic force microscopy, nanoparticle tracking have been

used for obtaining detailed phenotypic information but not for bulk sample processing, (Dragovic *et al.*, 2011; van der Pol E *et al.*, 2014). The different methods for MP quantification have been summarized in Fig. 3.

Microparticle Assessment by Flow Cytometry (FC)

As majority of the cell-derived MPs express PS on their surface, it becomes very easy to detect MPs by flow cytometer. Beads are used to make the gate which includes events of the size 1µm and below. After which these MPs can be tagged with fluorescent conjugated annexin V along with fluorescent conjugated antibodies for specific cell antigens to enumerate and characterize the cellular origin of the MPs. Flow Cytometer is available in most of the research institutes and many samples can be processed at one go. It is reported that the technique has an intra-assay and inter-assay variability of approximately 2-6% and 7-12% respectively (Shet *et al.*, 2003; Simak *et al.*, 2006; Brown *et al.*, 2011).

There had been many debates and concerns regarding the standardization of the FC technique and its pre-analytical variables. To overcome these problems, a first collaborative workshop to standardize PMP assessment using FC was initiated 6 years ago (Lacroix *et al.*, 2010a). The strategy involved the use of forward light scatter (FSC) signal of Megamix beads, Biocytex, which consist of 0.5, 0.9 and 3 µm beads so as to gate MPs such a manner that only those events 1 µm in size and below are included. Our center has participated in this workshop and standardized the technique on BD FACS Aria. However, it was observed that a better resolution and a more comparable response in SSC oriented instruments was obtained using side scatter signal (SSC) at 90°C, rather than FSC. The new beads called Megamix SSC beads were then used to make the gates for MPs in such instruments. A new workshop was, thus, initiated by International Society on Thrombosis and Haemostasis (ISTH) Vascular Biology Standardization Subcommittee to evaluate the inter-instrument variability (Coite *et al.*, 2017). This

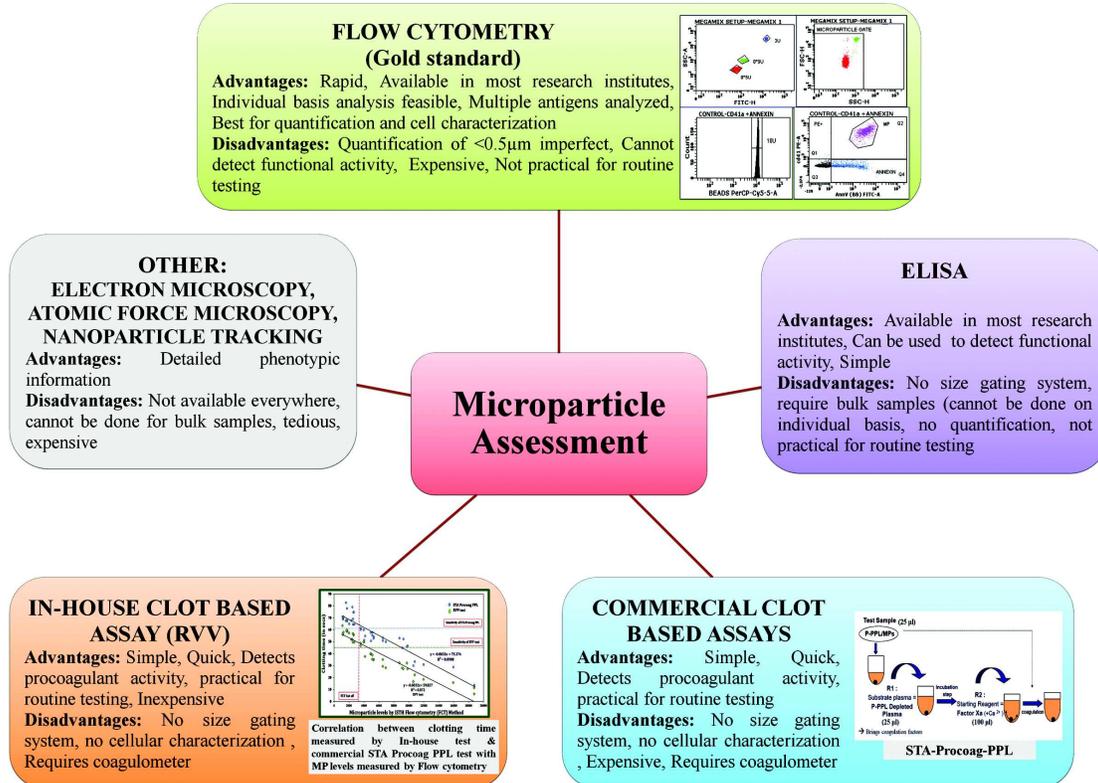


Fig. 3: Pros and Cons of different techniques for microparticle assessment: Flow cytometry is the gold standard method for MP analysis with the major limitation being that it cannot detect functional activity. Thus this method should always be complemented with a functional assay. Other methods used for MP detection are ELISA, clot based assays, and microscopy

assessment technique for MP analysis using FC is the most reliable method known. One important aspect of this method is the pre-analysis variables and processing which need to be kept uniform (Lacroix *et al.*, 2010b). The disadvantages of the technique are: (a) it works out to be an expensive test and though doable it is not practical for small number of samples to be processed on a daily basis for immediate results and (b) the technique cannot detect functional activity and thus needs to be performed in combination with functional assays.

Enzyme-linked Immune Sorbent Assay (ELISA)

In this technique, annexin V is coated in the well which captures PS expressing MPs after which origin of the MPs is detected by using a 2nd antibody specific to surface antigens of cell type. Different capture-antibodies and detection- antibodies have been used. This can further be used as a functional assay to check the prothrombinase activity of these MPs by first capturing them and then incubating them with a mixture of factor Xa, factor Va, prothrombin and calcium chloride after which a chromogenic substrate for thrombin is added. The advantages of this technique are that it is simpler, doable in maximum research facilities and allows the detection of procoagulant activity of MPs. The results have been found to correlate with those obtained by FC (Osumi *et al.*, 2001; Hugel *et al.*, 2004; Perez-Casal *et al.*, 2005). However, the major drawback is that as there is no gating system therefore, the size of what is being detected is unknown. Further, there is no insight into the size of MPs being analyzed. Other drawbacks are that the technique requires bulk sample processing rather than individual samples and the functional activity i.e. the prothrombinase activity cannot be assigned completely to MPs; other factors in the plasma may contribute to the same.

Clot Based Assays

A commercially available clot based assay for MPs i.e. STA Procoag PPL (DiagnosticaStago, France) is an efficient screening technique where in the results correlate well with FC. The method detects procoagulant activity on a semi-automated coagulometer based on the principle that procoagulant MPs will shorten the activated factor X clotting time. It is simple, quick, but expensive. Thus, we aimed to standardize an in-house clot based screening assay

for MP detection at our center which would not only be efficient but also inexpensive. In this assay, we measure the clotting time using semi-automated coagulometer after the addition of calcium chloride to MP rich plasma and incubating with Russell viper venom (RVV) and MP poor plasma. RVV activates factor X, thus triggering the coagulation cascade downstream of factor Xa. The MP poor plasma acts like a MP deficient plasma making the test dependent on procoagulant phospholipids on MPs in the samples. The results obtained from this test significantly correlated with the results obtained by FC ($R^2=0.87$, $p<0.01$). Thus, this assay can be utilized routinely for procoagulant MP assessment in various clinical settings. If the clotting time is shortened, FC test can be used to get exact levels and cell origin of MPs (Patil *et al.*, 2016a).

The most important aspect to remember is that irrespective of the technique used for MP assessment, the pre-analytical handling of MPs remains the most important cause of variability in the results. Therefore, the standardized methods need to be strictly adhered to (Vila-Liante *et al.*, 2016; Lacroix *et al.*, 2012).

Potential Role of Cell-derived MPs in Pregnancy Complications

MPs and Healthy Pregnancy

Normal pregnancy itself is known to be a hypercoagulable state marked with an increase in the procoagulant activity with elevated levels of factor VII, X, VIII, fibrinogen and vWF; and a decrease in anticoagulants with a major reduction in protein S activity and by acquired activated protein C (APC) resistance. Thus, it is well-known that pregnancy itself as a condition is an acquired risk for thrombosis (Greer, 1999; Heit *et al.*, 2001; Alijotas-Reig *et al.*, 2005). This phenomenon may protect the women from fatal hemorrhage during delivery but may also predispose her to thromboembolism.

In healthy pregnancy, studies suggest that the total platelet, endothelial, leukocyte and TF- bearing MPs as well as the procoagulant activity in healthy non-complicated pregnancy were higher in the 1st trimester as compared to non-pregnant age matched women; these MPs gradually increase during pregnancy with the highest levels observed in the 3rd trimester (Alijotas-Reig *et al.*, 2012; Radu *et al.*,

2015). Thus, increased MP levels may be one of the contributing factors to the hypercoagulable state seen in healthy pregnancy. There are other reports in which levels of MPs between normal pregnancies and non-pregnant controls do not differ suggesting that MPs are not related to pregnancy. However, MPs associated with fetal-derived human leukocyte antigen were significantly higher in normal pregnant women as compared to non-pregnant controls (Orozco *et al.*, 2009). Lok *et al.* (2008) observed reduced MPs which subsequently get normalized to the post-partum levels; but placenta-derived MPs were found to have increased.

MPs and Recurrent Pregnancy Loss (RPL)

Pregnancy loss (PL) is one of the most common complications of pregnancy which affects around 15% of reproducing couples and recurs in 2-3% of them. Despite a wide range of investigations, no apparent cause has been found in large number of cases (Coulam, 1991). RPL is defined as 2 or more failed pregnancy, wherein pregnancy is defined as a clinically documented pregnancy by ultrasonography or histopathological test (ASRM, 2013). One hypothesis that exists is that probably a defective maternal hemostatic response may lead to hypoxia and thrombosis of the uteroplacental vasculature and thus assist in subsequent fetal loss. The mechanisms which may be involved in this are impairment of trophoblast invasion, placental microthrombi and villitis (Greer, 1999; Rai, 2003).

Hereditary thrombophilia and antiphospholipid antibodies have found to have a strong association with PL and other pregnancy complications (Vora *et al.*, 2008; Hansda and Roychowdhury, 2012; Ocak *et al.*, 2013; Parand *et al.*, 2013; Chen *et al.*, 2015; Subrt *et al.*, 2013; Ching *et al.*, 2013). For this reason, genetic thrombophilia like protein C, protein S and antithrombin deficiency, factor V Leiden mutation; and antiphospholipid antibodies like lupus anticoagulant, anti-cardiolipin antibodies, anti- β_2 -glycoprotein-1 antibodies are being tested for in these patients. In a study conducted at our center (Patil *et al.*, 2015a), 587 women with no apparent etiological causes of RPL and 115 healthy women controls were tested for genetic thrombophilia markers and antiphospholipid antibodies. Among genetic thrombophilia, the risk of PL was highest in women

with protein S deficiency followed by plasminogen activator inhibitor-1 4G/4G polymorphism. Among antiphospholipid antibodies, the risk of PL was the highest in women with anti-cardiolipin antibodies, followed by anti-annexin antibodies and lupus anticoagulants. Thus, thrombophilia, both inherited and acquired, is an important contributing factor in unexplained RPL and should be screened.

In the past few years, cell-derived MPs have been found elevated in several prothrombotic conditions and may also be involved in the pathogenesis of RPL. As many cases still remain idiopathic even after being tested for the common thrombophilia markers, MPs seem to be the most plausible candidate for assisting RPL and other pregnancy complications like preeclampsia. Studies show a significant association between circulating EMPs and preeclampsia severity in combination with other markers. Thus, MPs in combination with other markers (serum, etc.) could be used as a good predictor in the 10-14th gestation weeks and to discriminate preeclampsia from other pregnancy complications (VanWijk *et al.*, 2002a, b; González-Quintero *et al.*, 2004; Marques *et al.*, 2012; Jadli *et al.*, 2017).

Majority of the reports have found elevated levels of MP during RPL. Table 1 summarizes reports on cell-derived MPs and recurrent or unexplained PL. Laude *et al.* (2001) studied 74 women with RPL, 49 being early PL (before 10th gestation week (GW)) and 25 being late PL (beyond 10th GW) and found that the MP's prothrombotic activity (tested by using prothrombinase assay) is much higher in women with RPL when compared to non-pregnant healthy controls.

Using the technique of FC, Carp *et al.* (2004) and Pasquier *et al.* (2013) showed that EMPs increase in women with recurrent miscarriage (RM) and unexplained PL. Kaptan *et al.* (2008) studied 20 women with RM and showed that PMP levels were significantly higher while the platelet activation marker, P-selectin had marginally increased. In a study done at our center, we found PS expressing TF and EMPs had significantly increased in women with RPL using FC technique and this corresponded with the procoagulant activity of MPs tested by the commercial kit STA Procoag PPL (Patil *et al.*, 2013). Another study (Martinez-Zamora *et al.*, 2016) found significantly higher levels of MPs in women with

antiphospholipid syndrome (APS) and also in women with unexplained fetal loss when compared to healthy controls; however, there was no statistical difference between these two groups. Contrary to these results, Toth *et al.* (2008) studied 51 women with RM and did not find significant difference between the test cases and controls with regard to total number of MPs or EMPs or PMPs. Alijotas-Reig *et al.* (2011) found a significant decrease in EMPs in PL group and RM group; the reason for decrease may be related to its consumption in clotting activation and thus removal from peripheral circulation. The most important point to note in all these studies is the time at which the sample was assessed for MPs. In the study conducted by Alijotas-Reig *et al.* (2011) blood samples were collected at the time of diagnosis of the loss and were compared to non-matched healthy pregnant controls. In the above mentioned studies including ours wherein elevated MP levels were observed, the blood samples were collected 4 months after the loss or delivery. This time was chosen as haemostatic changes noted during pregnancy normalize after delivery within 4 to 6 weeks whereas platelet count and protein S takes a little longer to normalize (Hellgren, 2003). In our study, we also found that % platelet MPs increase and become normal after anticoagulant therapy was initiated in pregnant women with a history of RPL. Thus % platelet MPs along with PS expressing MPs may be used to assess, diagnose and predict the pregnancy outcome. A decrease in % platelet MPs may suggest their consumption in fibrin deposit (Patil *et al.*, 2016b).

Thus, these studies show that presence of elevated endothelial, TF and PS expressing MPs suggest a continuous chronic endothelial damage or activation which may eventually get exaggerated at the onset of pregnancy. The data further corroborates that MPs may contribute and assist in uteroplacental thrombosis.

Therefore, in search for an underlying cause of unexplained RPL, focusing on circulating procoagulant MPs seems to be a promising approach. The above mentioned studies clearly show the presence of elevated procoagulant MPs in peripheral circulation in women with early and/or late unexplained/ RPL, thus adding to the new emerging body of evidence that MPs play a significant role in thrombosis-complicated disorders.

What are the Clinical Implications of Elevated MPs in RPL?

The next logical question is that if MPs are associated with PL by assisting in developing a prothrombotic state, what clinical approach would be beneficial for these patients? Will ACT which has proven to be beneficial in antiphospholipid syndrome and genetic thrombophilia help prevent thrombosis and thus subsequent PL in patients with elevated levels of procoagulant MPs?

The most debatable topic is whether all women with thrombophilia or unexplained PL need to be anticoagulated during pregnancy. There are contradictory reports with regards to the effect of ACT in pregnancy in women with thrombophilia. Some reports found that the occurrence of PL or pregnancy complications does not decrease with ACT (Rodger *et al.*, 2014; Schleussner *et al.*, 2015) whereas other studies show that ACT such as low dose aspirin, effective and preventive dose of heparin, low or immunosuppressive doses of corticosteroids, etc. are effective in reducing the incidence of adverse pregnancy outcomes in women with thrombophilia or history of idiopathic PL (Rosove *et al.*, 1990; Ghosh K *et al.*, 2008; Mitixæ *et al.*, 2011; Mutlu *et al.*, 2015).

There are no studies world-wide except a preliminary study done at our center with regards to assessing whether ACT would prove to be beneficial in patients with unexplained/RPL. Bulut *et al.* (2011) reported that aspirin reduces circulating EMPs and PMPs in patients with cardiovascular disease. In the study done at our center (Patil *et al.*, 2015b), we observed that procoagulant MPs are exaggerated at the onset of pregnancy; in 70% of these patients exaggerated MP levels decreased significantly as ACT progressed and normalized by the 3rd trimester. These women wherein the MPs normalized had successful live births; possibly as a result of ACT. These patients were on heparin and aspirin. The exact mechanism by which heparin and aspirin reduce the MP levels was not studied. However it is known that heparin has many attributes like being an anticoagulant, anti-inflammatory, inhibitory to complement activation, role in embryonic development, wound healing, metastasis, tissue homeostasis, cell differentiation and proliferation. Aspirin, on the other hand, is an antiplatelet agent that induces platelet aggregation and is a potent vasoconstrictor. Thus, these properties may

Table 1: Cell-derived microparticles and recurrent pregnancy loss

S.No.	Author	Patients and controls	Results	Method for MP detection	Time of blood sample collection
1	Laude I <i>et al.</i> , 2001	Patients: 74 women with unexplained PL: 49 early PL (before 10 th GW) 25 late PL (beyond 10 th GW) Controls: 50 non-pregnant women	MP's prothrombotic activity higher in women with RM when compared to non-pregnant healthy controls	Prothrombinase ELISA assay	At least 2 months from last obstetric event
2	Carp H <i>et al.</i> , 2004	Patients: 96 women with RPL Controls: 90 parous women	Elevated endothelial MPs were seen in a proportion of women with RPL (12.5%) when compared controls (2.2%) suggesting endothelial damage or activation	Flow cytometry using fluorescent anti-CD51/CD31 antibodies	3-4 months after previous miscarriage
3	Kaptan <i>et al.</i> , 2008	Patients: 20 women with RSA Controls: 20 age matched women	Platelet MPs were elevated in patients when compared to controls suggesting that PMPs may have a role in pathogenesis of RSA	Flow cytometry using platelet P-selectin (CD62P) as a platelet activation marker and CD42b(+)	At least 4 months from last obstetric event
4	Toth <i>et al.</i> , 2008	Patients: 51 women with RSA Controls: 24 non-pregnant parous women	Total annexin V binding MPs were similar in patients and controls. Elevated levels were observed in 10 patients when compared to 1 in controls. PMP, EMP, thrombin generation and F1+2 did not differ significantly suggesting no direct association between MPs and systemic coagulation activation in RSA patients	Flow cytometry using annexin V, and antibodies against CD61, CD63 and CD62P (PMP), as well as CD144 and CD62E (EMP). Prothrombin fragment 1 + 2 (F1+2) and thrombin generation were determined to assess the pro-coagulant potential of MP	At least 2 months from last obstetric event
5	Alijotas-Reig J <i>et al.</i> , 2011	Patients: 53 women with PL:30 RM16 UFL7 RM + UFLControls: 38 healthy pregnant women 20 healthy non-pregnant women controls	Significantly decreased endothelial MP levels were observed in patients with PL when compared to healthy pregnant controls and also in patients with RM when compared to healthy non-pregnant and CD41 MP/ μ L of plasma	Flow cytometry analyzing total annexin (A5+), endothelial-(CD144+/CD31+ CD41-), platelet-(CD41+), leukocyte-(CD45+)	At the time clinical diagnosis of miscarriage or fetal loss was made and after the patient was included into the study
6	Pasquier E <i>et al.</i> , 2013	Patients: 124 women with unexplained PLControls: 273 parous women	Higher endothelial MPs but lower platelet MPs in patients than controls suggesting endothelial damage	Flow cytometry using CD51, CD144, or CD146 for endothelial, CD41 for platelet, CD45 and CD66b for leukocyte and neutrophil MPs	At least 2 months from last obstetric event
7	Patil <i>et al.</i> , 2013	Patients: 115 women with RPL Controls: 20 healthy non-pregnant women	Total annexin V, TF and endothelial MPs were found significantly increased in patients when compared to controls. The clot time was also shortened in these samples showing increased procoagulant activity	Flow cytometry (standardized by participating in the ISTH Vascular Biology SSC workshop) analyzing total annexinV, platelet(CD41a), endothelial (CD146,CD62e), leukocyte (CD45), erythrocyte (CD235a) and tissue factor (CD142) (TF) expressing MPs and STA	At least 3 months from last obstetric event

8	Patil <i>et al.</i> , 2015b	Patients: 25 women on ACT with history of PL positive for > 1 thrombophilia but negative for other presumptive aetiological causes. Controls: 25 healthy age matched pregnant women with atleast one live birth and no history of thrombosis	In 15 of the 20 patients who showed exaggerated MPs at the onset of pregnancy, MP levels significantly reduced with ACT as pregnancy progressed and in 14 of them the MP normalized by the 3 rd trimester; all of them having successful pregnancy outcome	Procoag PPL commercial Clot test Flow cytometry (standardized by participating in the ISTH Vascular Biology SSC workshop) analyzing total annexinV MPs	In each trimester and 4 months after pregnancy outcome
9	Martinez-Zamora <i>et al.</i> , 2016	Patients APS group: 50 women with primary APS with >3 consecutive 1 st trimester miscarriages. UFL 52 women with with >3 consecutive 1 st trimester miscarriages of unknown etiology. Control: 52 healthy fertile women with no history of pregnancy loss	Cell- derived MPs were found significantly higher in women with APS and with UFL when compared to the control group. However group: there was no statistically significant difference UFL group derived MPs	Commerical functional assay- ZYMUPHEN MP- Activity, Hyphen BioMed was used to asses procoagulant cell	At least 6 months after last miscarriage
10	Patil <i>et al.</i> , 2016b	Patients:14 women with unexplained PL on ACT3 women with unexplained PL not on ACTControls: 25 healthy pregnant women	%PMPs is decreased at the beginning in women with unexplained PL but after ACT is initiated, the % increases and normalizes. %PMPs is highly decreased as pregnancy progresses in women not on therapy with the outcome being PL. %PMP rather than the PMP levels seem to be a clinically interesting marker along with PS expressing MPs to assess, diagnose and predict pregnancy outcome. A decrease in the %PMPs may suggest their consumption in fibrin deposits	Flow cytometry (standardized by participating in the ISTH Vascular Biology SSC workshop) analyzing total annexinV, platelet (CD41a) and endothelial(CD62e) MPs	In each trimester and 4 months after pregnancy outcome

MP: microparticles; GW: gestation week; PL: pregnancy loss; RM: recurrent miscarriage; RPL: recurrent pregnancy loss; RSA: recurrent spontaneous abortion; PMPs: platelet microparticles; EMP: endothelial microparticles; UFL: unexplained fetal loss; TF: tissue factor; ACT: anticoagulant therapy; APS: Antiphospholipid syndrome; ISTH: International Society of Thrombosis and Haemostasis; SSC: Scientific and standardization committee; STA Procoag PPL kit: Diagnostica Stago, France

be assisting in reducing the damage and activation of MP production and eventually resulting in a better outcome. In this study we also reported 2 patients who were only on Aspirin, wherein the exaggerated MPs at the start of pregnancy elevated even further as pregnancy progressed and both suffered from PL. Thus, aspirin alone may not be sufficient.

More studies are required to explain the essentiality of mechanism of reduction. In the search for diagnosis and treatment of unexplained RPL, these results seem promising. MPs may serve as important biomarkers for monitoring ACT.

Industrial Outlook

MPs play an important role in different clinical settings and also play a significant role in different biological processes like intercellular communication, coagulation, apoptosis and so on. In women with RPL, cell-derived procoagulant MPs were found to be statistically increased suggesting a chronic endothelial damage or activation which may eventually get exaggerated at the onset of pregnancy that may contribute and assist in uteroplacental thrombosis (Laude *et al.*, 2001; Carp *et al.*, 2004; Pasquier *et al.*, 2013;

Kaptan *et al.*, 2008; Patil *et al.*, 2013). Similarly MP concentrations are elevated in patients with cardiovascular disease (Baron *et al.*, 2012); MPs are found to be associated with type 2 diabetes which may result in cardiovascular complications (Nomura *et al.*, 2004). Hyper coagulation observed with cancer/malignancy has been attributed partly to procoagulant MPs (Furie *et al.*, 2005; Kim *et al.*, 2003). Thus, as MPs actively participate in different pathologies, modulation of MP release or clearance may be a novel pharmacological strategy for the management of the disease. In our study, we found exaggerated levels of MPs decreasing as pregnancy progresses in RPL patients on heparin along with aspirin; which resulted in successful pregnancy outcome. However, how heparin and aspirin help in reducing the MP levels is unknown (Patil *et al.*, 2015b). Thus MPs could be potential therapeutic targets and such inhibitors to MPs need to be identified.

Few of the currently identified inhibitors (Baron *et al.*, 2012; Roseblade *et al.*, 2013) include

- a) An increase in intracellular calcium plays a key role in MP generation; thus targeting the calcium influx can be one strategy to decrease MP production. However in different studies wherein calcium channel blockers were used, the outcome was positive but the MP levels were still much higher than healthy controls.
- b) Targeting calpain is one strategy due to its critical role in release of MPs.
- c) Clinical studies have shown the benefit of statins in cardiovascular diseases not only by reducing cholesterol, but possibly due to other modes like endothelial function, vascular inflammation, platelet aggregation, etc. Studies show that statins also reduce MP levels. The level of MPs were found reduced by anti-oxidant treatment reflecting a reduction in platelet activation as well as improvement in endothelial function.
- d) Anti-platelet drugs have been used for treatment of hypercoagulability in patients with diabetes mellitus. However, just like calcium channel blockers, the drugs are unable to reduce the MP concentration as compared to that of healthy controls.

MPs being a promising biomarker in different

clinical settings have even resulted in the standardization of different tests for their analysis like ELISAs, STA Procoag PPL, and other clot based simple screening assay. Novel diagnostic assays for analyzing different cell-derived MPs which are simple, cost effective as well as sensitive are the need of the hour. Thus, industries can initiate and produce novel methods, reagents and kits for analysis of MPs which can help in different branches of medicine.

Due to its ubiquitous role in multitude of clinical settings, there is a wide scope for industries to target this area; however being a new marker with a lot of unresolved issues, there is a need for more research to resolve the unanswered issues in the area of MPs.

Conclusions

The cell-derived MPs which were considered as futile and insignificant cellular debris in the earlier days are now emerging as novel important biomarkers of endothelial injury, inflammation and thrombosis. Apart from this, they have been found to play a crucial role in cellular communication, thus impacting numerous pathophysiological processes. The strong association with different disorders makes them very important clinical entities with regard to both diagnosis and treatment. However, research on MPs faces many problems such as methodology and pre-analytical variables and thus strict guidelines need to be adhered to for MP estimation. MPs will have true clinical utility as biomarkers of pathology for different disorders only when the methodology and pre-analytical handling of samples have been further standardized along with validation of the methods. Elevated procoagulant MPs have been detected in patients with pregnancy complication and unexplained RPL. This association of elevated procoagulant MPs in pregnancy complications has caught more attention as pregnancy itself being a hypercoagulable state, the presence of thrombophilia markers and elevated MPs may assist in uteroplacental thrombosis that leads to pregnancy complications and loss. Further, a preliminary study reports the reduction in elevated MP levels in women with PL as ACT progresses, where in these patients had successful live birth; highlighting the beneficial use of ACT in such patients with elevated MPs. Future studies may help explain the mechanism of action of ACT in reducing the MP levels. These findings may then be extended to other thrombosis complicated

diseases wherein MPs have been found elevated. Clearly, a lot of work needs to be done in this emerging and rising field of research and our understanding of these MPs will definitely expand significantly in future.

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